

1 ThruTracker: Open-Source Software for 2-D and 3-D Animal Video

2 Tracking

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12 Running headline: Open-source Animal Tracking Software

13 **Abstract**

- 14 1. Tracking animal movement patterns using videography is an important tool in many biological
15 disciplines ranging from biomechanics to conservation. Reduced costs of technology such as
16 thermal videography and unmanned aerial vehicles has made video-based animal tracking more
17 accessible, however existing software for processing acquired video limits the application of
18 these methods.
- 19 2. Here, we present a novel software program for high-throughput 2-D and 3-D animal tracking.
20 ThruTracker provides tools to allow video tracking under a variety of conditions with minimal
21 technical expertise or coding background and without the need for paid licenses. Notable
22 capabilities include calibrating the intrinsic properties of thermal cameras; tracking and counting
23 hundreds of animals at a time; and the ability to make 3-D calibrations without dedicated
24 calibration objects. Automated 2-D and 3-D workflows are integrated to allow for analysis of
25 largescale datasets.
- 26 3. We tested ThruTracker with two case studies. The 2-D workflow is demonstrated by counting
27 bats emerging from bridges and caves using thermal Videography. Tests show that ThruTracker
28 has a similar accuracy compared to humans under a variety of conditions. The 3-D workflow is
29 shown for making accurate calibrations for tracking bats and birds at wind turbines using only
30 the wind turbine itself as a calibration object.
- 31 4. ThruTracker is a robust software program for tracking moving animals in 2-D and 3-D. Major
32 applications include counting animals such as bats, birds, and fish that form large aggregations,
33 and documenting movement trajectories over medium spatial scales ($\sim 100,000 \text{ m}^3$). When
34 combined with emerging technologies, we expect videographic techniques to continue to see
35 widespread adoption for an increasing range of biological applications.

- 36 **Key words:** Animal Flight, Animal Movement, Movement Ecology, Population Monitoring, Flight
- 37 Biomechanics, Video Detection

38 1. Introduction

39 Video-based animal tracking is a widely used tool in fields as diverse as biomechanics, animal behavior,
40 ecology, and population monitoring (Dell et al., 2014). The reduced price of thermal video cameras,
41 high-speed cameras, and other technologies such as unmanned aerial vehicles (UAVs) has expanded
42 dramatically the capabilities of investigators studying animal movement in the lab and in the field
43 (Jackson, Evangelista, Ray, & Hedrick, 2016). This has placed great demand for video processing
44 software.

45 Various video tracking tools are available for two-dimensional (2-D), and three-dimensional (3-D) animal
46 tracking. Two-dimensional tracking is often used for studies of one or multiple individuals under
47 controlled laboratory settings. Two-dimensional tracking can also be useful for surveying animal
48 populations such as bats or birds and for monitoring their migratory behavior (Kunz et al., 2009).
49 Algorithms differ in how they detect animals and how they connect detections between frames to form
50 spatiotemporal tracks. For example, some software uses background subtraction for detection
51 (Rodriguez et al., 2018) while other programs use adaptive thresholding (Sridhar et al., 2019). One
52 popular program uses image recognition to help track individuals (Pérez-Escudero et al. 2014). Deep
53 learning based approaches have recently become popular, particularly for marker-less tracking of animal
54 body parts (Mathis et al., 2018; Pereira et al., 2019).

55 Three-dimensional animal tracking uses two or more cameras to triangulate animal positions. This
56 requires synchronized video acquisition, careful calibration of the optical properties of cameras
57 [i.e. camera extrinsics). Several software programs are available for generating 3-D calibrations and
58 tracking animals in 3-D (Noldus et al., 2001; Hedrick, 2008; Theriault et al., 2014; Jackson et al., 2016;
59 Knorlein et al., 2016; Nath et al., 2019).

60 Originally, 3-D calibrations required that an object with several markers at known 3-D positions be
61 placed within the field of view of each camera (Abdel-Aziz & Karara, 1971). More recent workflows
62 produce calibrations by moving an object with a recognizable 2-D pattern [“checkerboard calibration”
63 (Zhang, 2000)] or with two markers at a fixed distance (“wand calibration”) through the calibration
64 volume (Theriault et al., 2014). This allows one to set the scale of the scene using the wand length or
65 pattern scale and to estimate the accuracy of the calibration based on the variation of reconstructed
66 wand lengths. However, researchers are increasingly interested in studying animals within large volumes
67 in the natural environment where it is difficult to deploy calibration objects or where doing so might
68 disturb the animals under study (Evangelista, Ray, Raja, & Hedrick, 2017; Corcoran & Hedrick, 2019). In
69 addition, much of the currently available 3-D tracking software requires manual digitization of objects in
70 the videos, which limits the amount of data that can be processed. Finally, calibration procedures are
71 not widely available for determining the intrinsic properties of thermal cameras, preventing their
72 widespread use for 3-D tracking applications.

73 Here we present ThruTracker, a free and open-source software package for high-throughput 2-D and 3-
74 D animal tracking. ThruTracker provides an app-based environment (i.e., no coding required) with all the
75 tools necessary to track animals under a variety of conditions with light-based or thermal cameras.
76 ThruTracker is coded in MATLAB v2020b (Natick, MA, USA) and compiled versions of the software are
77 provided so that it can be installed and run for free without any licenses. We provide source code under
78 a 3-clause BSD license for those who may want to customize the software for their own purposes.
79 ThruTracker is also compatible with any tracking software that outputs 2-D coordinates, and it exports
80 data in standard text formats that can be imported into other software for further processing or
81 statistical analysis.

82 We first provide an overview of the 2-D and 3-D track generation process, before demonstrating the use
83 of ThruTracker with two test cases—tracking bats at wind turbines and counting bat emergences from

84 bridges and caves using a thermal camera flown with an Unmanned Aerial Vehicle (UAV) or recording
85 from the ground. We conclude by discussing applications of 3-D tracking and current limitations. A step-
86 by-step manual for ThruTracker along with additional recommendations for camera setups and use of
87 the software is available on our website (www.sonarjamming.com/thrutracker).

88 2. Materials and Methods

89 2.1 Workflow for 2-D and 3-D Tracking

90 ThruTracker can be used either for 2-D or 3-D video tracking. Two-dimensional tracking does not require
91 that the videos be calibrated and is therefore a much simpler procedure. Three-dimensional tracking
92 requires calibration of the camera properties such as focal length and lens distortion (intrinsic
93 calibration) and determination of the positions and orientations of the cameras for each scene where
94 they are deployed (extrinsic calibration). The workflow for 3-D track generation is shown in Figure 1.

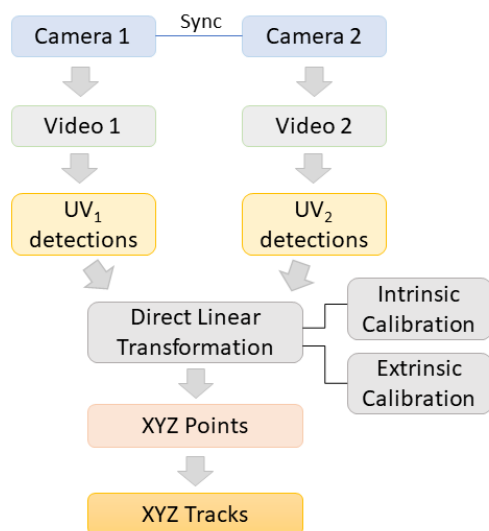


Figure 1. Overview of 3-D track generation procedure. Two or more cameras acquire synchronized video, which are each processed to generate 2-D (UV) detections. Intrinsic and extrinsic camera calibrations are used to generate direct linear transformation (DLT) equations that transform UV coordinates into real-world 3-D or XYZ points for each detection. XYZ points are then stitched together across frames to generate 3-D tracks.

95
96 Step 1: Video Acquisition. Synchronized videos are acquired using two or more cameras. Some cameras,
97 such as FLIR A65 thermal video cameras (FLIR Systems, Inc., Wilsonville, OR) used in our testing, have
98 dedicated electronic inputs that allow digital signals to synchronize the shutters of each camera. An

99 alternative for cameras without sync ports is to use audio signals that are broadcast to each camera
100 (Jackson et al., 2016). Audio synchronization of cameras does not allow for shutter synchronization,
101 therefore there will be a time offset of up to half the frame rate even after synchronization. Recording
102 at 30 Hz, a half-frame synchronization accuracy has been sufficient to achieve highly accurate
103 calibrations for animals moving at low to moderate pixel speeds (Corcoran & Hedrick, 2019). Audio
104 synchronization may also require fine-tuning since some camera models may not keep the audio and
105 video outputs in perfect alignment.

106 Step 2: Object detection. After videos are acquired, videos from each camera are processed to detect
107 moving objects in each frame. ThruTracker uses a Gaussian mixture-based background subtraction
108 algorithm implemented in OpenCV [“BackgroundSubtractorMOG2”, (Zivkovic, 2004; Zivkovic & Van Der
109 Heijden, 2006)]. A blob detector is then used to isolate detections and the blob centroids are used as
110 detection coordinates. A simplified interface allows the user to rapidly select and modify detection
111 settings for their application. Settings include: 1) selecting which frames to process, 2) minimum and
112 maximum object size in pixels, 3) sensitivity for adjusting the threshold for discriminating foreground
113 from background, 4) Number of background frames used for generating the rolling background image,
114 and 5) target object diameter. This last option determines the size of a 2-D gaussian filter that is applied
115 to the image, which helps reduce noise and isolate closely spaced animals. Each frame of the video is
116 processed for detections before detections are linked together into 2-D or 3-D tracks in the proceeding
117 steps. Two-dimensional tracking applications can skip to step 6.

118 Step 3: Intrinsic camera calibration. For 3-D tracking applications, one must determine the camera’s
119 intrinsic properties, including the focal length, principal point and lens distortion (Abdel-Aziz & Karara,
120 1971; Hartley & Zisserman, 2004; Lourakis & Argyros, 2009). This information can be used to map any
121 pixel in the camera image into a vector in a camera-based frame. Typically, this calibration can be done
122 once per camera and lens combination in a laboratory and the values should be similar between

123 cameras and lens of the same make and model. In some special circumstances one might need to
124 calibrate each individual camera-lens pair for increased accuracy. Cameras with variable focal length
125 (i.e., a zoom lens) typically need to be calibrated separately at each focal length used for recording.

126 We use MATLAB's built-in camera calibration functions (Bouguet, 1999) with some modifications to
127 calibrate thermal images (Yahyanejad, Misiorny, & Rinner, 2011). We recommend using MATLAB's built-
128 in camera calibrator app or freely available functions in OpenCV or Argus (Jackson et al., 2016) for
129 calibrating light-based cameras.

130 Traditionally, calibration procedures for light-based cameras rely on detecting the corners of a
131 checkerboard pattern. With thermal imaging, one must first heat the checkerboard so that the darker
132 squares will be hotter than the lighter squares because of higher light absorption (Figure 2a). For
133 example, we achieved this by moving two 100-watt lamps over the checkerboard pattern for about 20
134 seconds before taking thermal images.

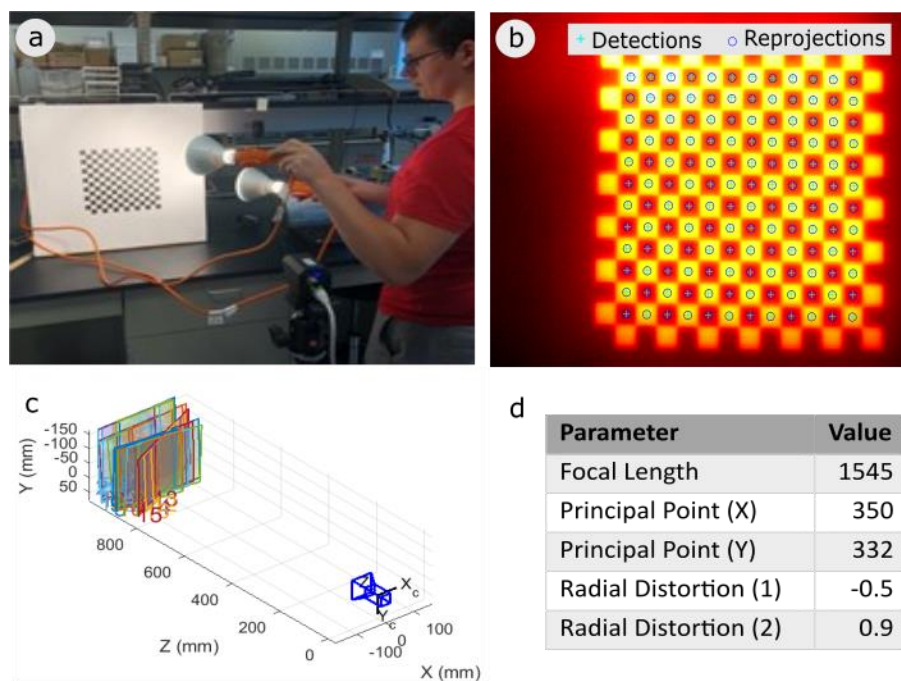


Figure 2. Thermal camera intrinsic calibration. (a) An experimenter heats a checkerboard pattern with two 100-watt lamps. (b) Example thermal image of a checkerboard pattern with detected and reprojected square centers. (c) Graphical depiction of the camera and 28 images of the checkerboard pattern taken at different positions and orientations. (d) Example output of intrinsic camera parameters for a Flir A65 camera.

135

136 However, uneven heating combined with rapid conduction of heat through the checkerboard image

137 increases the difficulty of detecting the corners between light and dark squares (or in this case between
138 cold and hot squares; Figure 2b). Instead of detecting the corners, the cooler portions of the image are
139 dilated so that each hot square is reduced in size and no longer connected to adjacent hot squares. A
140 blob-detector is then used to detect the centers of the hot squares. The pixel values of the image are
141 then inverted, and the procedure is repeated to detect the centers of the cold squares. This is repeated
142 for 20-30 images taken at different positions and orientations relative to the camera. The resulting data
143 are used by the software to calculate the camera's intrinsic parameters (Figure 2c, d).

144 Step 4: Extrinsic camera calibration. A second calibration procedure is required for generating 3-D tracks
145 for each recording setup—that is, any time the cameras are moved even slightly. The extrinsic
146 calibration determines the positions of the cameras in real-world 3-D space and their orientations [roll,
147 yaw and pitch; (Abdel-Aziz & Karara, 1971)]. Together with the intrinsic calibration data, the extrinsic
148 calibration allows one to map objects that are detected in two or more cameras to real-world 3-D
149 coordinates.

150 As noted previously, there are several approaches for generating extrinsic calibrations. ThruTracker has
151 two options: a wand-based procedure that is based on methods used for easyWand (Lourakis & Argyros,
152 2009; Theriault et al., 2014) and a procedure that is based exclusively on background points. One can
153 also import calibrations made using DLTdv or easyWand (Hedrick, 2008; Theriault et al., 2014).
154 Background points include any object that is visible within both cameras such as tips of tree branches,
155 wind turbines, or animals. Moving objects can be used if the images are taken synchronously between
156 cameras. This background point procedure [also known as structure from motion (Schönberger &
157 Frahm, 2016)] means that 3-D calibrations can be made without dedicated calibration objects within the
158 field of view. In effect, any object visible by two or more cameras can be used as a calibration object.

159 Two additional components of the calibration must be determined when no dedicated calibration
160 objects are used: the scale of the scene and the gravitational axis. The scale can be set by specifying two
161 points in the scene that are at a known distance from one another. Alternatively, one can use the
162 distance between cameras to set the camera scale.

163 Second, the gravitational axis can be set by measuring the inclination angle of one of the cameras using
164 an inclinometer. This value is input into ThruTracker's calibration app during the calibration procedure.

165 With these options, one can obtain calibration data rapidly in the field with only a few measurements
166 and with no requirement for deploying calibration objects. This is especially helpful for situations where
167 it is not feasible to deploy calibration objects or where their use would disturb animals under study.

168 A notable down-side of not using calibration objects is the absence of objects at known positions that
169 can be used to check the accuracy of the calibration. Therefore, it is important to conduct tests of the
170 calibration procedure using objects at known distances. For example, in our test using wind turbines
171 below, we measured the variation in the reconstructed lengths of the turbine blades at different times
172 and positions throughout our recording. Another alternative would be to conduct test setups in the field
173 at locations where it is easier to deploy calibration objects such as a wand.

174 Step 5: Generating 3-D Points. Three-dimensional points are generated after objects have been detected
175 in videos from each camera and intrinsic and extrinsic calibrations are completed. For each set of
176 synchronized video frames, theoretical 3-D points are created from all combinations of 2-D detections
177 across cameras. Each putative 3-D point has an associated direct linear transformation (DLT) residual.
178 The DLT residual is the distance in pixels between the observed image coordinates of a marker and the
179 "ideal" image coordinates computed from the estimated 3-D location of the marker and the calibration
180 information for the camera that captured the image. If we visualize each 2-D detection as a vector in 3-D
181 space with its origin at the camera, the residuals indicate how closely a given set of 2-D detections and

182 their associated vectors come to crossing in 3-D space. The algorithm starts by creating 3-D points based
183 on sets of 2-D points with the lowest residuals and removing those points from the available pool. It
184 proceeds until there are no more 3-D points with residuals below the specified threshold.

185 Step 6: Generating 2-D and 3-D Tracks. Two-dimensional and Three-dimensional points are stitched
186 together across frames to make tracks. Each 2-D or 3-D point in the first frame is a putative track.
187 Detections in each proceeding frame are assigned to existing tracks, or if no assignment is made, they
188 become the beginning of a new track. A Kalman filter is applied to each track to predict the position of
189 the track in the next frame. The distance between each detection and the predicted positions are
190 computed. This is done both in 2-D and in 3-D to calculate a cost matrix. A Hungarian algorithm is used
191 to determine the combination of assignments that minimizes cost across the assignment matrix (Kuhn,
192 1955). Finally, a threshold is specified in ThruTracker such that assignments are only made if their cost is
193 below the threshold. One can specify the number of frames between detections that are allowed (i.e.,
194 gap distance) before a track is terminated. One also specifies the minimum number of detections
195 required for a track to be retained. Longer gaps and smaller minimum numbers of detections increase
196 the number of tracks that are retained but increases the number of false positive tracks generated from
197 noise.

198 Step 7: Data visualization, analysis and classification: ThruTracker offers multiple tools for visualizing and
199 processing tracks. One can rapidly toggle between tracks and use shortcut keys to classify them into
200 different categories. For example, for wind turbine applications, the tracks can be labeled “bird”, “bat”,
201 “airplane”, “noise”, etc. Another option allows all the tracks to be visualized at once. Tracks can then be
202 selected as groups and classified based on their positions, start or end points. This tool is helpful for
203 selecting tracks based on their location, as exhibited in the bat emergence case study presented below.
204 Another option allows the user to draw a rectangle over the camera image to count exits and re-entries
205 as animals pass into or out of the rectangle. This is useful, for example, when counting bats exiting a

206 cave roost. Resulting 2-D, 3-D and track data can be exported into CSV files for use in other analysis
207 programs.

208 2.2 Case Studies

209 2.2.1 Case Study 1: Counting bat exits from bridges and caves using thermal imaging.

210 We used ThruTracker to count bats emerging from bridges using a DJI Zenmuse XT2 thermal camera
211 with a 13 mm lens (45-degree field of view) suspended from a DJI Matrice 300 drone (SZ DJI Technology
212 Co., Shenzhen, China). Because this analysis was done in 2-D there was no need for intrinsic or extrinsic
213 calibrations. The drone was flown at altitudes of 50 m and 80 m above a bridge known to be a roost
214 location for big brown bats (*Eptesicus fuscus*) in August of 2020 near Burnsville, NC, USA. We also
215 counted gray bats leaving caves using thermal cameras placed at ground level. Videos of gray bats were
216 provided by the US Fish and Wildlife Service.

217 Our goals were 1) to determine the maximum distance at which bats could be detected and 2) to
218 compare manual counts of emergences with those produced using ThruTracker. The bridge recordings
219 provide a test of relatively low numbers of bats counted near the limits of their detection range. The
220 cave recordings test detection of large numbers with high rates of occlusion.

221 Videos were processed in ThruTracker with the following parameters: sensitivity, 35; background
222 frames, 20; Min object pixels, 1; max object pixels 100; min track length 5, max gap length 5; match
223 threshold 10. After detections were made in ThruTracker, the applet TrackSelector was used to rapidly
224 select tracks that originated near the edge of the bridge. Manual observers used VirtualDub software to
225 play videos at 50% of normal speed and paused and reviewed videos frame by frame, as necessary.

226 We processed two videos taken at heights of 50 m and 80 m above the bridge and two videos
227 representing different bat densities at caves (Table 1). Videos were not meant to census the entire
228 emergences, but rather provide data for comparing detection abilities.

229 2.2.2 Case Study 2:

230 We tested ThruTracker's ability to make 3-D calibrations for tracking 3-D flights of bats and birds at wind
231 turbines. Studying animal movements at wind turbines is a problem of considerable interest, especially
232 for bats who are being killed in large numbers for mostly unknown reasons (Arnett & Baerwald, 2013).
233 Thermal imaging has been used for studying bats at wind turbines in 2-D (Horn, Arnett, & Kunz, 2008;
234 Cryan et al., 2014) and 3-D (Kinzie et al., 2018; Schirmacher, 2020). Proprietary software for 3-D tracking
235 of bats at wind turbines (was also recently published (Matzner, Warfel, & Hull, 2020). Here we report on
236 our field camera setup, intrinsic and extrinsic calibration. Because we conducted these tests under cold
237 winter conditions with few bats or birds present, we did not obtain 3-D animal tracks. For example
238 tracks using a similar approach, see Schirmacher, 2020 (Schirmacher, 2020).

239 We tested the calibration setup using two Flir A65 thermal cameras with 25-degree lenses synchronized
240 with electronic inputs at an experimental wind turbine at the National Renewable Energy Laboratory
241 (NREL) in Golden, CO during December 2019. The cameras were placed 33 m apart from one another
242 and 40 m from the base of the turbine's monopole. Cameras were aimed slightly below the wind
243 turbine's nacelle, which was 80 m off the ground. This resulted in an inclination angle of 62.2 degrees for
244 our reference camera.

245 3. Results

246 3.1 Case Study 1: Counting bat exits from bridges and caves using thermal imaging.

247 ThruTracker detected similar numbers of bats compared to the manual observers in all test cases, with
248 error rates of 0.6-7.1% (Table 1; Figure 1; Supplemental Video 1. Thermal video taken from a UAV
249 showed that the 50 m recording height was comfortably within the detection range for the bats. Bat
250 detections had a size of 5.2 ± 3.4 pixels (mean \pm st. dev.) and tracks extended across most of the image
251 (Figure 3a) including over water and over the bridge. However, bats failed to be tracked over some land

252 areas where they lacked contrast with the background (e.g., see tracks terminating as they approach
253 land area in bottom right portion of Figure 3a). Manual inspection of videos at these locations found
254 that bats were not readily visible to the human eye, so this appears to be a limitation of the thermal
255 imaging, not the detection algorithm.

256 The 80 m recording height was much closer to the detection limit for *E. fuscus* bats using this recording
257 setup. Bats had a detection size of 1.9 ± 1.1 pixels. Visual inspection of videos revealed that bats were
258 only barely visible and that they were not detectable to the human eye within a short distance of their
259 emergence. This is reflected in the ThruTracker tracks that terminate over open water some distance
260 from the bridge exits (Figure 3b). This may have resulted from bats dropping to a lower altitude (and
261 further distance from the camera) as they left the bridge, as was visible in other camera views taken at
262 an oblique angle relative to the ground. ThruTracker performed slightly worse under these at the 80 m
263 height than the 50 m height, but still detected 93% of tracks that were detected manually (Table 1).

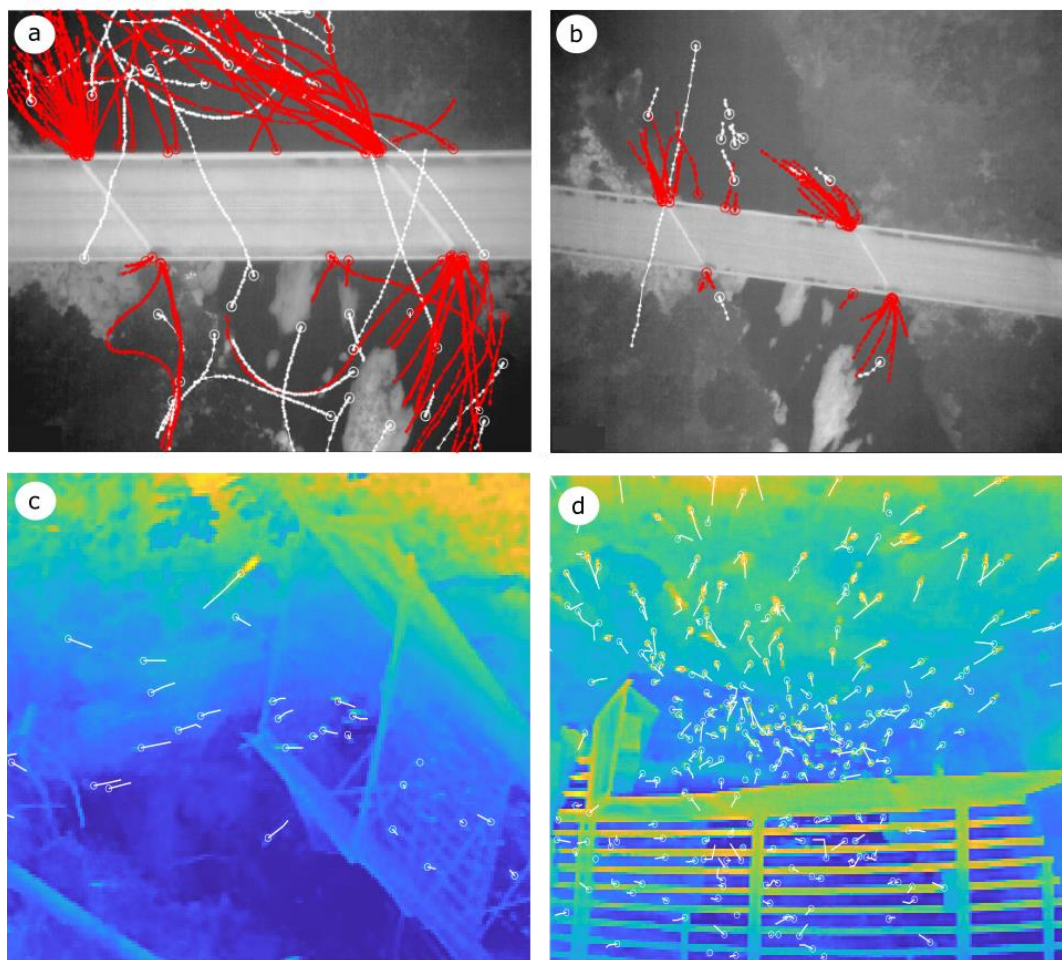
264 Counts of gray bats (*Myotis grisescens*) exiting caves were used to test of ThruTracker's multi-object
265 tracking abilities. ThruTracker achieved error rates of 4.3% and 0.6% for the two test videos, which we
266 estimated to have exit rates of 16.4 and 103.0 bats per second (Table 1; Figure 3c, d; Supplemental
267 Video 2).

268 *Table 1. Comparison of manual and automated bat counts using thermal videography*

	Video 1	Video 2	Video 3	Video 4
Emergence type	Bridge	Bridge	Cave	Cave
Camera height (m)	50	80	ground	ground
Duration (m:ss)	3:21	1:58	0:05	0:05
ThruTracker detections	86	39	85	518
Manual detections*	85.5 (85,86)	42 (42,42)	81.5 (81, 82)	515 (508,522)
Exit rate (bats per second)	0.42	0.35	16.4	103.0
Error rate	0.6%	7.1%	4.3%	0.6%

269 *Values show means of 2 observers; values in parentheses show values of the two observers.

Figure 3 Example ThruTracker detections of bats leaving a bridge (a, b) and cave (c, d). Big brown bats (*Eptesicus fuscus*) were filmed exiting a bridge using a thermal camera on a UAV flying at 50 m (a) and 80 m altitude (b). Red tracks indicate exits and white tracks indicate other detections. In (a, b) circles indicate the starting point of tracks to highlight departures from the bridge and the entirety of all tracks are shown. In (c, d) detections from a single frame are shown (circles) with tracks indicating movement over the two previous frames. See table 1 for statistics.



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272 3.2 Case Study 2: Calibrating large 3-D volumes using wind turbines.

273 In our second case study, we demonstrate ThruTracker's 3-D workflow for calibrating large spatial

274 volumes using only the turbine itself as a calibration object. We calibrated the FLIR A65 thermal cameras

275 (640 x 512 pixel resolution) using the intrinsic calibration method described above. Example images and
276 resulting intrinsic camera parameters are shown in Figure 2. We used 28 checkerboard images, with an
277 average reprojection residual error of 0.69 pixels (range 0.48-0.83 pixels).

278 We generated an extrinsic calibration in ThruTracker using 67 points from the wind turbine as
279 background points (Figure 3). These included hot spots, corners and an anemometer on the nacelle, and
280 turbine blade tips. Points were digitized manually using DLTdv8 (Hedrick, 2008). Efforts were made to
281 select points that were 1) clearly visibly in both cameras, 2) distinct points in 3-D space, such as small
282 hot spots or sharp edges of objects, and 3) covering a broad range of 2-D and 3-D positions. We
283 excluded six points because they had DLT residuals > 3 pixels. The remaining calibration had mean
284 reprojection errors of 0.63 pixels.

285 We set the scale of the scene using the distance between the two cameras (33 m) and the gravitational
286 axis was set using the inclination angle of the second camera (62.2 degrees). The resulting calibration
287 had a volume of 235,597 m³ assuming a maximum detection distance of 200 m. The maximum detection
288 range is likely less than 200 m for small bats (<30 g) with this camera setup, but it is possible that some
289 large birds could be detected at this distance.

290 To test the spatial accuracy of our calibration, we measured the distance between the tips of turbine
291 blades and the tip of the hub in 23 frames chosen to represent a variety of spatial configurations. This
292 resulted in a mean distance of 41.4 m, standard deviation of 1.05 m, and coefficient of variation of 0.02.
293 Therefore, we can expect typical errors less than ± 1 -2 m for this calibration setup. For comparison, a
294 recently published study testing similar software for tracking birds and bats at wind turbines found
295 errors up to ± 20 m (Matzner et al., 2020).

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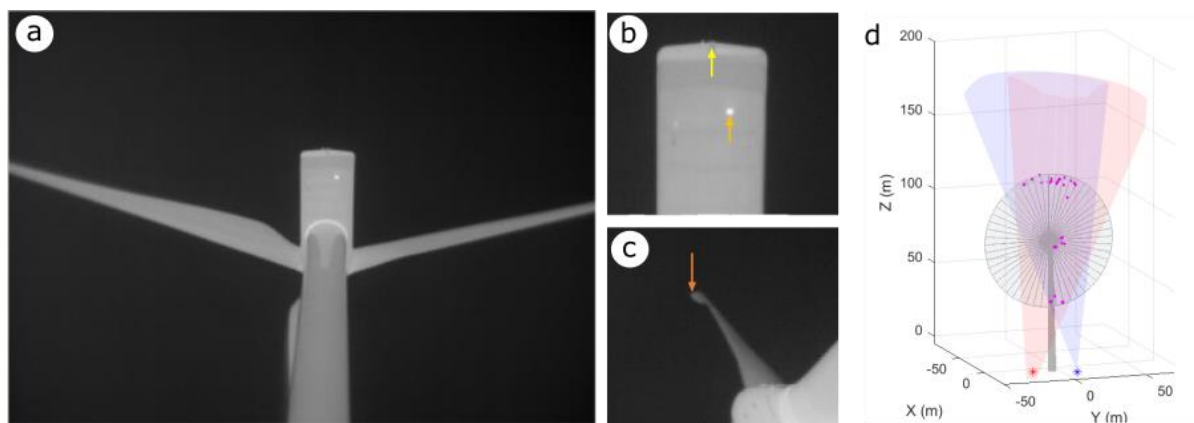


Figure 4. Extrinsic calibration of a wind turbine scene. (a) Example views from a single camera. (b, c) Examples of background points used for generating the calibration. These include a back corner of the nacelle (yellow arrow), a small hotspot of unknown origin (orange arrow) and a turbine tip (dark orange arrow). (d) Visualization of the calibrated scene. Views of two cameras are shown in light red and light blue shading. The wind turbine is shown as a grey outline but note that the turbine rotates about the vertical axis depending on wind direction. Magenta points show 3-D positions of points used for making the calibration.

297

298 4. Discussion

299 4.1. Animal Tracking Applications

300 Videographic techniques are seeing expanded use for studies of wildlife (Cilulko, Janiszewski,
301 Bogdaszewski, & Szczygielska, 2013; Christiansen et al., 2014; Gonzalez et al., 2016) ranging from
302 animals in agricultural fields (Christiansen et al., 2014), to cetaceans (Seymour, Dale, Hammill, Halpin, &
303 Johnston, 2017), bats and birds (Betke et al., 2008; Cullinan, Matzner, & Duberstein, 2015; Matzner,
304 Cullinan, & Duberstein, 2015) and ants (Narendra & Ramirez-Esquivel, 2017). One major application of
305 videography for studies of animals in the natural environment is counting populations. For example, 3-D
306 thermal imaging has been used to show that bat colonies have only a small fraction of the number of
307 individuals compared to earlier human counts (Betke et al., 2008). Thermal imaging has also been used
308 in combination with radar and acoustics for monitoring migratory patterns of birds (Gauthreaux &
309 Livingston, 2006; Horton, Shriver, & Buler, 2015).

310 A second major application of videography is studying animal movement patterns. Numerous studies
311 have investigated the structure and rules underlying bird flocks (Ballerini et al., 2008; Evangelista et al.,
312 2017) and schools of fish (Jolles et al., 2017). Videography has also been used for studying flight
313 biomechanics of animals under natural conditions et al., 2015), and interactions between bats and birds
314 and large structures such as wind turbines and oil and gas platforms (Horn et al., 2008; Cryan et al.,
315 2014; Ronconi et al., 2015). Most of the studies described above have relied on custom software that is
316 not widely available. The aim of the current study was to develop a robust, easy to use and free
317 software package that could be used for these and other applications.

318 4.2 ThruTracker Capabilities

319 Here we present a new software package for 2-D and 3-D animal tracking. ThruTracker has several
320 features not found in other freeware. These include easily adjustable procedures for 2-D and 3-D
321 tracking; a tool for calibrating intrinsic parameters of thermal cameras; the ability to track and count
322 hundreds of animals simultaneously; and the ability to make 3-D calibrations without dedicated
323 calibration objects. We demonstrate these capabilities by counting bats leaving bridges and caves (Table
324 1; Figure 3) and making a 3-D calibration using only a wind turbine as a calibration object (Figure 4).

325 The software is compatible with thermal and light-based imaging and most standard video formats (e.g.,
326 avi, wmv, mp4). ThruTracker uses an app-based environment with no coding required to make well-
327 established detection and tracking algorithms widely available. Users simply import videos and select
328 the detection and tracking options. These features make it easy to track moving animals under a variety
329 of conditions.

330 4.3 Requirements and Limitations

331 ThruTracker uses a well-established background subtraction algorithm for object detection (Zivkovic,
332 2004; Zivkovic & Van Der Heijden, 2006). This method generates a rolling model of the background using

333 a number of frames that can be specified by the user. This approach is best suited for stationary
334 backgrounds and animals that are in near continuous motion. It has difficulty with animals that stay in
335 one place; however, using more images for generating the background would help address this problem.
336 ThruTracker aims to detect one point for each animal (the blob centroid). For detecting multiple body
337 parts per animal one should consider deep-learning based approaches (Mathis et al., 2018; Pereira et al.,
338 2019). ThruTracker has built-in compatibility for importing detections from other programs such as
339 DeepLabCut (Mathis et al., 2018) for could be used for generating 3-D tracks from point clouds.

340 The main requirements for tracking in 3-D are 1) synchronized video are acquired from at least two fixed
341 cameras 2) camera intrinsics are calibrated in the laboratory, and 3) some objects (including the focal
342 animals themselves) are visible at a range of 2-D and 3-D positions within the calibrated volume.

343 Without the requirements for dedicated calibration objects, it is now possible to calibrate nearly any
344 volume in the lab or field. We demonstrate this workflow for generating 3-D calibrations at wind
345 turbines, where it would be logistically challenging to put calibration objects in the airspace (Figure 4).
346 Another approach would be to use the animals themselves as background points (Corcoran & Hedrick,
347 2019).

348 5. Conclusions

349 Technological development is driving price reductions and capability expansion in thermal and high-
350 speed cameras, along with supporting equipment such as UAVs. However, the software required to
351 make full use of these capabilities for research in fields as diverse as biomechanics, animal behavior,
352 ecology, and population monitoring remains the province of specialized workflows in individual lab
353 groups. ThruTracker provides an integrated, graphical, and user-friendly package to fill these needs, thus
354 expanding the number of researchers able to make effective use of these emerging technologies.

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364 Authors' Contributions

365 A.C. and T.H conceived of and developed the software. M.S. provided funding and guided software
366 development. E.B. collected data used for testing and aided with data analysis. A.C. drafted the
367 manuscript and all authors edited the manuscript and gave final approval for the publication.

368 Data Availability Statement

369 ThruTracker installation files, a user manual and software tutorials are available at
370 www.sonarjamming.com/thruTracker. Source code is available as a GitHub repository
371 (<https://github.com/AaronJCorcoran/ThruTracker>).

372 References

- 373 Abdel-Aziz, Y. I., & Karara, H. M. (1971). Direct linear transformation from comparator coordinates into
374 object space coordinates in close-range photogrammetry. In *Proceedings of the Symposium on*
375 *Close Range Photogrammetry* (pp. 1–18). American Society for Photogrammetry and Remote
376 Sensing. doi:10.14358/PERS.81.2.103
- 377 Arnett, E. B., & Baerwald, E. F. (2013). Impacts of Wind Energy Development on Bats: Implications for
378 Conservation. In R. Adams & S. Pederson (Eds.), *Bat Evolution, Ecology, and Conservation* (pp. 435–
379 456). New York: Springer. doi:10.1007/978-1-4614-7397-8
- 380 Ballerini, M., Cabibbo, N., Candelier, R., Cavagna, A., Cisbani, E., Giardina, I., ... Zdravkovic, V. (2008).
381 Interaction ruling animal collective behavior depends on topological rather than metric distance:
382 Evidence from a field study. *Proceedings of the National Academy of Sciences of the United States*
383 *of America*, 105(4), 1232–1237. doi:10.1073/pnas.0711437105
- 384 Betke, M., Hirsh, D. E., Makris, N. C., McCracken, G. F., Procopio, M., Hristov, N. I., ... Others. (2008).
385 Thermal imaging reveals significantly smaller Brazilian free-tailed bat colonies than previously
386 estimated. *Journal of Mammalogy*, 89(1), 18–24.
- 387 Bouguet, J.-Y. (1999). *Visual methods for three-dimensional modeling*. Cal Tech. *PhD Dissertation*
388 <https://resolver.caltech.edu/CaltechETD:etd-02072008-115723>
- 389 Christiansen, P., Steen, K., Jørgensen, R., & Karstoft, H. (2014). Automated Detection and Recognition of
390 Wildlife Using Thermal Cameras. *Sensors*, 14(8), 13778–13793. doi:10.3390/s140813778
- 391 Cilulko, J., Janiszewski, P., Bogdaszewski, M., & Szczygielska, E. (2013, February). Infrared thermal
392 imaging in studies of wild animals. *European Journal of Wildlife Research*. Springer Verlag.
393 doi:10.1007/s10344-012-0688-1

- 394 Corcoran, A. J., & Hedrick, T. L. (2019). Compound-v formations in shorebird flocks. *ELife*, 8.
395 doi:10.7554/eLife.45071
- 396 Cryan, P. M., Gorresen, P. M., Hein, C. D., Schirmacher, M. R., Diehl, R. H., Huso, M. M., ... Dalton, D. C.
397 (2014). Behavior of bats at wind turbines. *Proceedings of the National Academy of Sciences of the*
398 *United States of America*, 111(42), 15126–31. doi:10.1073/pnas.1406672111
- 399 Cullinan, V. I., Matzner, S., & Duberstein, C. A. (2015). Classification of birds and bats using flight tracks.
400 *Ecological Informatics*, 27, 55–63. doi:10.1016/j.ecoinf.2015.03.004
- 401 Dell, A. I., Bender, J. A., Branson, K., Couzin, I. D., de Polavieja, G. G., Noldus, L. P. J. J., ... Brose, U. (2014,
402 July). Automated image-based tracking and its application in ecology. *Trends in Ecology and*
403 *Evolution*. Elsevier Ltd. doi:10.1016/j.tree.2014.05.004
- 404 Evangelista, D. J., Ray, D. D., Raja, S. K., & Hedrick, T. L. (2017). Three-dimensional trajectories and
405 network analyses of group behaviour within chimney swift flocks during approaches to the roost.
406 *Proceedings of the Royal Society B. Biological Sciences*, 284(1849), 20162602.
407 doi:10.1098/rspb.2016.2602
- 408 Gauthreaux, S. A., & Livingston, J. W. (2006). Monitoring bird migration with a fixed-beam radar and a
409 thermal-imaging camera. *Journal of Field Ornithology*, 77(3), 319–328. doi:10.1111/j.1557-
410 9263.2006.00060.x
- 411 Gonzalez, L., Montes, G., Puig, E., Johnson, S., Mengersen, K., & Gaston, K. (2016). Unmanned aerial
412 vehicles (UAVs) and artificial intelligence revolutionizing wildlife monitoring and conservation.
413 *Sensors*, 16(1), 97. doi:10.3390/s16010097
- 414 Hartley, R., & Zisserman, A. (2004). *Multiple View Geometry in Computer Vision*. Cambridge University
415 Press. doi:10.1017/cbo9780511811685

- 416 Hedrick, T. L. (2008). Software techniques for two- and three-dimensional kinematic measurements of
417 biological and biomimetic systems. *Bioinspiration and Biomimetics*, 3(3), 6. doi:10.1088/1748-
418 3182/3/3/034001
- 419 Horn, J. W., Arnett, E. B., & Kunz, T. H. (2008). Behavioral Responses of Bats to Operating Wind Turbines.
420 *Journal of Wildlife Management*, 72(1), 123–132. doi:10.2193/2006-465
- 421 Horton, K. G., Shriver, W. G., & Buler, J. J. (2015). A comparison of traffic estimates of nocturnal flying
422 animals using radar, thermal imaging, and acoustic recording. *Ecological Applications*, 25(2), 390–
423 401. doi:10.1890/14-0279.1.sm
- 424 Jackson, B. E., Evangelista, D. J., Ray, D. D., & Hedrick, T. L. (2016). 3-D for the people: Multi-camera
425 motion capture in the field with consumer-grade cameras and open source software. *Biology*
426 *Open*, 5(9), 1334–1342. doi:10.1242/bio.018713
- 427 Jolles, J. W., Boogert, N. J., Sridhar, V. H., Couzin, I. D., & Manica, A. (2017). Consistent individual
428 differences drive collective behavior and group functioning of schooling fish. *Current Biology*,
429 27(18), 2862-2868.e7. doi:10.1016/j.cub.2017.08.004
- 430 Kinzie, K., Hale, A., Bennett, V., Romano, B., Skalski, J., Coppinger, K., & Miller, M. F. (2018). Ultrasonic
431 Bat Deterrent Technology. Technical Report DE-EE007035 doi:/10.2172/1484770
- 432 Knorlein, B. J., Baier, D. B., Gatesy, S. M., Laurence-Chasen, J. D., & Brainerd, E. L. (2016). Validation of
433 XMALab software for marker-based XROMM. *Journal of Experimental Biology*, 219(23), 3701–3711.
434 doi:10.1242/jeb.145383
- 435 Kuhn, H. W. (1955). The Hungarian method for the assignment problem. *Naval Research Logistics*
436 *Quarterly*, 2(1–2), 83–97. doi:10.1002/nav.3800020109
- 437 Kunz, T. H., Betke, M., Hristov, N. I., & Vonhof, M. J. (2009). Methods for assessing colony size, and

- 438 relative abundance of bats. In T. H. Kunz & S. Parsons (Eds.), *Ecological and Behavioural Methods*
439 *for the Study of Bats* (pp. 133–157). Baltimore, MD: Johns Hopkins University Press.
- 440 Lourakis, M. I. A., & Argyros, A. A. (2009). SBA: A software package for generic sparse bundle
441 adjustment. *ACM Transactions on Mathematical Software*, *36*(1), 1–30.
442 doi:10.1145/1486525.1486527
- 443 Mathis, A., Mamidanna, P., Cury, K. M., Abe, T., Murthy, V. N., Mathis, M. W., & Bethge, M. (2018).
444 DeepLabCut: markerless pose estimation of user-defined body parts with deep learning. *Nature*
445 *Neuroscience*, *21*(9), 1281–1289. doi:10.1038/s41593-018-0209-y
- 446 Matzner, S., Cullinan, V. I., & Duberstein, C. A. (2015). Two-dimensional thermal video analysis of
447 offshore bird and bat flight. *Ecological Informatics*, *30*, 20–28. doi:10.1016/J.ECOINF.2015.09.001
- 448 Matzner, S., Warfel, T., & Hull, R. (2020). ThermalTracker-3-D: A thermal stereo vision system for
449 quantifying bird and bat activity at offshore wind energy sites. *Ecological Informatics*, *57*, 101069.
450 doi:10.1016/J.ECOINF.2020.101069
- 451 Narendra, A., & Ramirez-Esquivel, F. (2017). Subtle changes in the landmark panorama disrupt visual
452 navigation in a nocturnal bull ant. *Philosophical Transactions of the Royal Society B: Biological*
453 *Sciences*, *372*(1717). doi:10.1098/rstb.2016.0068
- 454 Nath, T., Mathis, A., Chen, A. C., Patel, A., Bethge, M., & Mathis, M. W. (2019). Using DeepLabCut for 3-D
455 markerless pose estimation across species and behaviors. *Nature Protocols*, *14*(7), 2152–2176.
456 doi:10.1038/s41596-019-0176-0
- 457 Noldus, L. P. J. J., Spink, A. J., & Tegelenbosch, R. A. J. (2001). EthoVision: A versatile video tracking
458 system for automation of behavioral experiments. *Behavior Research Methods, Instruments, and*
459 *Computers*, *33*(3), 398–414. doi:10.3758/BF03195394

- 460 Pereira, T. D., Aldarondo, D. E., Willmore, L., Kislin, M., Wang, S. S. H., Murthy, M., & Shaevitz, J. W.
461 (2019). Fast animal pose estimation using deep neural networks. *Nature Methods*, *16*(1), 117–125.
462 doi:10.1038/s41592-018-0234-5
- 463 Pérez-Escudero, A., Vicente-Page, J., Hinz, R. C., Arganda, S., & De Polavieja, G. G. (2014). IdTracker:
464 Tracking individuals in a group by automatic identification of unmarked animals. *Nature Methods*,
465 *11*(7), 743–748. doi:10.1038/nmeth.2994
- 466 Rodriguez, A., Zhang, H., Klaminder, J., Brodin, T., Andersson, P. L., & Andersson, M. (2018). *ToxTrac* : A
467 fast and robust software for tracking organisms. *Methods in Ecology and Evolution*, *9*(3), 460–464.
468 doi:10.1111/2041-210X.12874
- 469 Ronconi, R. A., Allard, K. A., & Taylor, P. D. (2015, January 1). Bird interactions with offshore oil and gas
470 platforms: Review of impacts and monitoring techniques. *Journal of Environmental Management*.
471 Academic Press. doi:10.1016/j.jenvman.2014.07.031
- 472 Schirmacher, M. (2020). Evaluating the effectiveness of an ultrasonic acoustic deterrent in reducing bat
473 fatalities at wind energy facilities. Technical Report DOE-BCI-0007036
- 474 Schönberger, J. L., & Frahm, J.-M. (2016). Structure-from-Motion Revisited. In *Proceedings of the IEEE*
475 *Conference on Computer Vision and Pattern Recognition (CVPR)* (pp. 4104–4113).
- 476 Seymour, A. C., Dale, J., Hammill, M., Halpin, P. N., & Johnston, D. W. (2017). Automated detection and
477 enumeration of marine wildlife using unmanned aircraft systems (UAS) and thermal imagery.
478 *Scientific Reports*, *7*(1), 1–10. doi:10.1038/srep45127
- 479 Shelton, R. M., Jackson, B. E., & Hedrick, T. L. (2014). The mechanics and behavior of cliff swallows
480 during tandem flights. *Journal of Experimental Biology*, *217*(15).
- 481 Sholtis, K. M., Shelton, R. M., & Hedrick, T. L. (2015). Field Flight Dynamics of hummingbirds during

- 482 territory encroachment and defense. *PLOS ONE*, 10(6), e0125659.
- 483 doi:10.1371/journal.pone.0125659
- 484 Sridhar, V. H., Roche, D. G., & Gingsins, S. (2019). Tracktor: Image-based automated tracking of animal
485 movement and behaviour. *Methods in Ecology and Evolution*, 10(6), 815–820. doi:10.1111/2041-
486 210X.13166
- 487 Theriault, D. H., Fuller, N. W., Jackson, B. E., Bluhm, E., Evangelista, D., Wu, Z., ... Hedrick, T. L. (2014). A
488 protocol and calibration method for accurate multi-camera field videography. *Journal of*
489 *Experimental Biology*, 217(11).
- 490 Yahyanejad, S., Misiorny, J., & Rinner, B. (2011). Lens distortion correction for thermal cameras to
491 improve aerial imaging with small-scale UAVs. In *ROSE 2011 - IEEE International Symposium on*
492 *Robotic and Sensors Environments, Proceedings* (pp. 231–236). doi:10.1109/ROSE.2011.6058528
- 493 Zhang, Z. (2000). A flexible new technique for camera calibration. *IEEE Transactions on Pattern Analysis*
494 *and Machine Intelligence*, 22(11), 1330–1334. doi:10.1109/34.888718
- 495 Zivkovic, Z. (2004). Improved adaptive Gaussian mixture model for background subtraction. In
496 *Proceedings - International Conference on Pattern Recognition* (Vol. 2, pp. 28–31). Institute of
497 Electrical and Electronics Engineers Inc. doi:10.1109/icpr.2004.1333992
- 498 Zivkovic, Z., & Van Der Heijden, F. (2006). Efficient adaptive density estimation per image pixel for the
499 task of background subtraction. *Pattern Recognition Letters*, 27(7), 773–780.
- 500 doi:10.1016/j.patrec.2005.11.005
- 501